Review article

Design strategies and applications of nacre-based biomaterials

Ethan Michael Gerhard, Wei Wang, Caiyan Li, Jinshan Guo, Ibrahim Tarik Ozbolat, Kevin Michael Rahn, April Dawn Armstrong, Jingfen Xia, Guoying Qian, Jian Yang

Department of Biomedical Engineering, Materials Research Institute, The Huck Institutes of the Life Sciences, The Pennsylvania State University, University Park, PA 16802, USA
Zhejiang Provincial Top Key Discipline of Bioengineering, College of Biological and Environmental Sciences, Zhejiang Wanli University, South Qianhu Road 8, Ningbo 315100, China
Engineering Science and Mechanics Department, Materials Research Institute, The Huck Institutes of the Life Sciences, The Pennsylvania State University, University Park, PA 16802, USA
Department of Orthopaedics and Rehabilitation, College of Medicine, The Pennsylvania State University, Hershey, PA 17033, USA

Keywords:
Biomaterials
Nacre
Pearl powder
Pearl
Bone
Orthopedic
Tissue engineering

Abstract

The field of tissue engineering and regenerative medicine relies heavily on materials capable of implantation without significant foreign body reactions and with the ability to promote tissue differentiation and regeneration. The field of bone tissue engineering in particular requires materials capable of providing enhanced mechanical properties and promoting osteogenic cell lineage commitment. While bone repair has long relied almost exclusively on inorganic, calcium phosphate ceramics such as hydroxyapatite and their composites, the organically derived shell and pearl nacre generated by mollusks has emerged as a promising alternative. Nacre is a naturally occurring composite material composed of inorganic, calcium carbonate plates connected by a framework of organic molecules. Similar to mammalian bone, the highly organized microstructure of nacre endows the composite with superior mechanical properties while the organic phase contributes to significant bioactivity. Studies, both in vitro and in vivo, have demonstrated nacre's biocompatibility, biodegradability, and osteogenic potential, which are superior to pure inorganic minerals such as hydroxyapatite or non-degradable metals. Nacre can be used directly as a bulk implant or as part of a composite material when combined with polymers or other ceramics. While nacre has demonstrated its effectiveness in multiple cell culture and animal models, it remains a relatively underexplored biomaterial. This review introduces the formation, structure, and characteristics of nacre, and discusses the present and future uses of this biologically-derived material as a novel biomaterial for orthopedic and other tissue engineering applications.

Statement of Significance

Mussel derived nacre, a biological composite composed of mineralized calcium carbonate platelets and interplatelet protein components, has recently gained interest as a potential alternative ceramic material in orthopedic biomaterials, combining the integration and mechanical capabilities of calcium phosphates with increased bioactivity derived from proteins and biomolecules; however, there is limited awareness of this material's potential. Herein, we present, to our knowledge, the first comprehensive review of nacre as a biomaterial. Nacre is a highly promising yet overlooked biomaterial for orthopedic tissue engineering with great potential in a wide variety of material systems. It is our hope that publication of this article will lead to increased community awareness of the potential of nacre as a versatile, bioactive ceramic capable of improving bone tissue regeneration and will elicit increased research effort and innovation utilizing nacre.
1. Introduction

Nacre, commonly known as mother of pearl, is a naturally occurring composite material composed of aragonite (calcium carbonate) platelets cemented by a complex organic matrix. Nacre comprises the inner layer of pearl oyster and freshwater pearl mussel shells as well as the pearls themselves, with pearl and shell nacre demonstrating similar chemical compositions as well as high strength, mechanical resilience, and iridescence.

Previous research has demonstrated nacre's biocompatibility, biodegradability, and osteogenic properties, marking nacre's potential as an alternative to other commonly utilized biomaterials in tissue engineering. Nacre has been shown to facilitate osteoblast proliferation and accelerate extracellular matrix (ECM) production and mineralization [1-4]. Additionally, nacre has demonstrated high mechanical strength, excellent bioconductivity, osteogenic capability, and biodegradability both in vitro and in vivo. The organic matrix contained within nacre has also been found to contain biological molecules capable of activating osteoblasts through chemical signaling; however, research into these organic molecules has been limited [5,6].

Nacre is a composite consisting of an inorganic mineral phase and an organic matrix, similar to the native structure of bone. This organized structure imparts nacre with its remarkable mechanical strength, while the presence of the organic matrix imparts improved osteoconductivity compared to synthetic materials such as titanium [7]. While orthopedic tissue engineering has previously relied on either autografts or allografts (limited by tissue availability and donor site morbidity) or permanent metal implants, biodegradable materials and composites are the focus of much research in order to provide grafting materials with high availability as well as the potential to be replaced over time by native tissue [8,9]. Nacre has recently emerged as a potential bone grafting material due to the above-mentioned favorable properties. Bulk nacre as well as composite materials containing nacre as the mineral phase and utilizing various polymers including poly(ε-lactic acid) (PLLA), poly(ε,ε-Lactide) (PDLLA), poly(ε,ε-Lactide-co-glycolide) (PLGA), ultra-high molecular weight polyethylene (UHMWPE), Polyetheretherketone (PEEK), and poly(3-hydroxybutyrate-co-3-hydroxyvalerate) (PHBV) have been tested as potential biomaterials.

Despite its demonstrated biocompatibility and bioactive properties, to date, no comprehensive review of nacre based materials has been reported and the potential role of nacre based materials in tissue engineering still remains relatively unknown. This paper summarizes current research on nacre, including its basic characteristics as well as its potential use as a biomaterial. Through an examination of relevant literature, the potential of nacre as an alternative to commonly used compositing materials such as hydroxyapatite (HA) and other calcium phosphate minerals for bone tissue engineering will be expounded. Elaboration of the previously conducted research on nacre will demonstrate its potential as a novel and relatively unexplored material for orthopedic biomaterials science and will stimulate continued research into nacre as an alternative to commonly used biomaterial ceramics such as hydroxyapatite.

2. Characteristics of nacre

2.1. Formation of nacre

Nacre is an organized organic-inorganic composite material, secreted by the epithelial cells of the mantle tissue of mollusks. Mantle epithelial cells secrete crystal precursors (CaCO3 and CO32-) as well as amorphous calcium carbonate, along with organic matrix molecules consisting predominantly of proteins and polysaccharides [10]. In this manner, calcium carbonate forms between the outer layer of mantle epithelial cells and the periostracum membrane. The organic matrix is co-deposited along with the mineral phase, and serves as a critical mineralization regulator.

In nacre, sheets of beta-chitin are interspersed by a hydrophobic gel, containing acidic, aspartic acid rich proteins, forming a highly structured chitinous framework [11]. Amorphous calcium carbonate is deposited at crystal nucleation sites, where it acts as seed molecules for the growth of the aragonite platelets that make up the mineral phase of nacre [12]. The process of nacre formation can be summarized as: (1) the formation of a layer of elongated calcite prisms, arranged tightly in a perpendicular orientation to the direction of shell growth, followed by (2) the crystallization of calcium carbonate within the platelets, converting amorphous calcite to a layer of flat, crystalline aragonite platelets [11]. This process is conserved among many species of mollusk (see Table 1).

Pearl formation, when occurring naturally as opposed to artificial seeding, is characterized as a host immune response to intrusion of a foreign body. Typically, a foreign particle or parasite enters the mollusk, prompting local irritation of the mantle tissue and subsequent formation of a pearl sac to cover the site. A similar process occurs during the pearl forming process when a seed is implanted in the mantle tissue of the mollusk to stimulate the above foreign body response and generate a pearl. Once a pearl sac is formed, nacre is secreted layer by layer to envelope and isolate the irritant, resulting in formation of a pearl after several years. Nacreous pearls are primarily produced by molluscan bivalves or clams. Of the many other molluscan species, few can produce pearls of interest as gemstones. Pearl producing species include the bailer shell *Melo*, the giant clam, *Tridacna*, various scallop spe-
cies, pen shell *Pinna*, and the *Haliotis iris* species of abalone. Saltwater pearls can be grown in several species of marine pearl oysters in the family *Pteriidae*. Freshwater pearls grow in several species of freshwater mussels including the families *Unionidae* and *Margaritiferidae*.

### Table 1

Mollusk species capable of secreting nacre.

<table>
<thead>
<tr>
<th>Species</th>
<th>Ref.</th>
<th>Species</th>
<th>Ref.</th>
</tr>
</thead>
<tbody>
<tr>
<td>Bivalvia</td>
<td></td>
<td>Bivalvia</td>
<td></td>
</tr>
<tr>
<td><em>Unio pictorum</em></td>
<td>FM [17,18]</td>
<td><em>Pinctada martensi</em></td>
<td>MPM [19,20]</td>
</tr>
<tr>
<td><em>Chamberlainia hainesiana</em></td>
<td>FPM [21]</td>
<td><em>Pinctada maxima</em></td>
<td>MPM [22,23]</td>
</tr>
<tr>
<td><em>Hyriopsis desowitzi</em></td>
<td>FPM [21]</td>
<td><em>Pinna rudis</em></td>
<td>MPM [19]</td>
</tr>
<tr>
<td><em>Hyriopsis myersiana</em></td>
<td>FPM [21]</td>
<td><em>Pteria chinensis</em></td>
<td>MPM [29]</td>
</tr>
<tr>
<td><em>Lamelldens marginalis</em></td>
<td>FPM [30]</td>
<td><em>Pteria hirundo</em></td>
<td>MPM [31]</td>
</tr>
<tr>
<td><em>Margaritifera dahurica</em></td>
<td>FPM [33]</td>
<td><em>Gastropoda</em></td>
<td></td>
</tr>
<tr>
<td><em>Margaritifera dahurinaia</em></td>
<td>FPM [34,35]</td>
<td><em>Biomphalaria glabrata</em></td>
<td>FPM [36]</td>
</tr>
<tr>
<td><em>Margaritifera laevis</em></td>
<td>FPM [33]</td>
<td><em>Calliostoma zyzyphus</em></td>
<td>FPM [13]</td>
</tr>
<tr>
<td><em>Margaritifera margaritifera</em></td>
<td>FPM [37]</td>
<td><em>Gibbula pennant</em></td>
<td>MM [13]</td>
</tr>
<tr>
<td><em>Margaritifera middendorffi</em></td>
<td>FPM [33]</td>
<td><em>Gibbula umbilicalis</em></td>
<td>MM [13]</td>
</tr>
<tr>
<td><em>Unio tumidus</em></td>
<td>FPM [38]</td>
<td><em>Haliotis iris</em></td>
<td>MM</td>
</tr>
<tr>
<td><em>Cristaria plicata</em></td>
<td>FPM [39]</td>
<td><em>Haliotis laevigata</em></td>
<td>MM [40,41]</td>
</tr>
<tr>
<td><em>Lamprotula leai</em></td>
<td>FPM [42]</td>
<td><em>Haliotis rufescens</em></td>
<td>MM [43]</td>
</tr>
<tr>
<td><em>Bathymodiolus azorius</em></td>
<td>MM [44]</td>
<td><em>Monodonta sp. ME</em></td>
<td>MM [13]</td>
</tr>
<tr>
<td><em>Isognomon radiatus</em></td>
<td>MM [19]</td>
<td><em>Trochus niloticus</em></td>
<td>MM</td>
</tr>
<tr>
<td><em>Malleus albus</em></td>
<td>MM [19]</td>
<td><em>Turbo marmoratus</em></td>
<td>MM</td>
</tr>
<tr>
<td><em>Nucula sulcata</em></td>
<td>MM [13]</td>
<td><em>Cephalopoda</em></td>
<td></td>
</tr>
<tr>
<td><em>Pteria avicula</em></td>
<td>MM [19]</td>
<td><em>Nautilus macromphalus</em></td>
<td>MM [45,46]</td>
</tr>
<tr>
<td><em>Atrina pectinata</em></td>
<td>MM* [19]</td>
<td><em>Nautilus pompilius</em></td>
<td>MM [47]</td>
</tr>
<tr>
<td><em>Mytilus edulis</em></td>
<td>MPM [48]</td>
<td><em>Monoplacophorans</em></td>
<td></td>
</tr>
<tr>
<td><em>Pinctada fucata</em></td>
<td>MPM [49]</td>
<td><em>Veleropilina zografi</em></td>
<td>MM [14]</td>
</tr>
</tbody>
</table>

* FM, freshwater mollusks; FPM, Freshwater pearl mollusks; MPM, Marine pearl mollusks; MM, marine mollusks.

---

**Fig. 1.** Microstructure of nacre in *P. margaritifera*. (A) Opened shell displaying internal, iridescent shell nacre layer and brown mantle tissue. (B) Schematic representation of the shell structure and overlying mantle. (C) and (D) The “brick and mortar” structure of the nacreous layer. Reprinted from [46], with permission from Elsevier.
mortar' or 'brick wall' microstructure [46,47,51–53]. Within the ‘brick and mortar’ structure (Fig. 1), aragonite tablets and thin sheets of organic matrix are alternatively arranged, with aragonite tablets acting as the stacked ‘bricks’ and the organic macromolecules acting as the ‘mortar’, resulting in the superior mechanical properties of the nacre composite [46,54–56]. This characteristic microarchitecture imparts a fracture resistance over 3000 fold higher than bulk, geological aragonite [57].

Although nacre is formed in a tightly controlled process, there are several types including columnar and sheet structures. Columnar nacre consists of tablets arranged in a vertical orientation and stacked in columns across the layers and is typically found in Cephalopoda and Gastropoda. Sheet nacre, in contrast, consists of tablets deposited at the margins of the underlying tablet, resulting in a more random arrangement and is typically found in bivalve species such as oysters and mussels [58].

Nacre is formed from the accumulation of polygonal tablets of aragonite crystal. Typical crystal size ranges from 5 to 15 μm in diameter and 0.4 to 0.5 μm thick, separated by sheets of interlamellar organic matrixes [53,59–64]. Within the interlamellar organic matrixes, interlamellar sheets are well ordered in a parallel orientation and less than 80 nm from one another [28,60]. Wang revealed the presence of nanolaths in the nacre tablets about 100 nm in width, with large block twins dividing the nacre tablet into two sectors [52]. Further investigation into the crystallization mechanism of the single crystal nacre tablet could further elucidate the reason and importance of these features.

At the microscale, nanoasperites are present on the surface of the tablets, which have been proposed to act as sources of resistance to tablet sliding. Moreover, the interface between tablets shows a prominent waviness [65]. This waviness contrasts with the previously supposed smooth interface model. The presence of this microstructural motif is thought to be critical to the inelastic deformation and strength of nacre.

Nacreous tablets are stiff and have a high tensile strength, while the interface between tablets has a high compressive stiffness and strength. The hierarchically assembled structure of tablets and matrix endows the composite with its excellent tensile strength and toughness [65,66]. The mechanical properties of mollusk nacre and shell have been summarized by Heinemann [67]. Due to its structural arrangement, nacre exhibits outstanding mechanical properties: elastic modulus: 64.7 ± 3.50 GPa, density: 2.6 g/cm³, and hardness: 4.41 ± 0.45 GPa [68]. Jackson investigated the mechanical characteristics of nacre from the bivalve Pinctada, demonstrating a tensile strength of 140–170 MPa, Young’s modulus of 60–70 GPa, and three point bending value of 350–1240 J/m² depending on the hydration state [69]. Sarikaya reported wet nacre of the red abalone shell (H. rufescens) obtained a fracture toughness of 7 ± 3 MPa and a strength of 180 ± 20 MPa while wet nacre of P. margaritifae had a fracture toughness of 10 ± 6 MPa and a strength of 220 ± 60 MPa [70].

Inspired by the mechanical properties of naturally occurring nacre, much recent research has focused on nacre mimetic materials. Generally, such study has attempted to combine a ceramic component with a relatively elastic, polymeric material. Such combinations offer the distinct advantage of combining the high toughness and damage resistance of ceramic materials with the elasticity and deformability of polymers, resulting in materials with improved fracture resistance, toughness, and mechanical strength. Schmieden created bacteria generated nacre mimetic structures by alternating deposition of CaCO₃ by ureolytic bacteria and bacteria-ally produced γ-polyglutaminate [71]. Hu utilized liquid crystal self-templating (LCST) to fabricate nacre inspired materials, assembled in a mimetic hierarchy of alternating inorganic nanoplatelets and biomacromolecules. These nacre mimetic composites displayed tensile strengths five to eight times higher than that of natural nacre and excellent ductility (one to two orders of magnitude greater than nacre) [72]. Voet used poly (vinylidene fluoride) lamellae covered with poly (methacrylic acid) as a template to direct silica mineralization, resulting in organized, nacre-like layers [73]. Bai created poly (methyl methacrylate) (PMMA)/HA composites with nacre mimetic architecture using bidirectional freezing. The obtained materials exhibited moduli of up to 20 GPa, bending strengths up to 100 MPa, and a fracture strength of up to 2075 J/m² (~100x monolithic HA) [74]. Hao combined poly (amido amine) and clay nanosheets, cross-linked by genipin, creating biocompatible lamellar composites with a fracture toughness of up to 5103 MJ/m² [75]. Further modification revealed that smaller plates suffered from pull-out failure, while in larger plates, failure by fracture dominated, with the former modality displaying inelastic deformation and the latter toughness and strength similar to cross-linked materials [78].

Graphene has attracted much interest recently as a biomaterial due to its bioactivity, biocompatibility, electrical conductance, and potential for surface modification. As a tailorable nanomaterial, graphene is an ideal candidate for the ceramic phase in nacre mimetic materials. Wan fabricated strong, conductive nacre-like structures using vacuum assisted filtration to align graphene oxide (GO) within a chitosan matrix using the electrostatic interactions between the positively charged chitosan and negatively charged GO under alkaline conditions [79]. Fabricated structures displayed strength 4-10x natural nacre [79]. Lian utilized similar methods to create nacre-like structures from gelatin and ultra large GO, attaining tensile stresses of up to 630.4 MPa through annealing [80]. Duan also achieved high tensile strength and toughness using nanofibrillar cellulose and GO cross-linked with 10,12 pentacosadiyn-1-ol. The impressive mechanical properties were attributed to the breakage of hydrogen bonds followed by the chain stretching of the polymer phase and finally the breakage of covalent bonds, providing a large amount of mechanical dissipation [81]. Chen created paper-like composite films of sodium alginate and GO, with strengths of up to 200 MPa [82]. Li used carboxyl graphene to control the deposition of CaCO₃, generating nacre-like morphology with CaCO₃ concentration dependent structures [83]. Wan combined GO with molybdenum disulfide nanosheets and thermoplastic polyurethane to form high strength ternary composites with high toughness due to the crack bridging effect of molybdenum disulfide [84]. Yan generated multifunctional, magnetic, luminescent, and conductive composites combining Fe₃O₄ and lanthanide doped upconversion nanocrystals surface modified with positively charged poly (ethylene imine) and negatively charged GO [85]. Tan successfully fabricated organized structures combining GO and various polymer phases including chitosan, PVA, and poly (ethylene oxide) (PEO), utilizing the attractive forces between charged groups to overcome the repulsion of GO molecules to effect a gel-sol transition and fabricate materials with tensile strengths of up to 200 MPa [86]. Combining ceramics and polymeric phases, nacre-mimetic materials with excellent mechanical properties and multiple functional properties including electrical conductance, fluorescence and optical transparency can be
obtained. Given the high degree of modification inherent in this type of assembly as well as the potential of such materials to elucidate composite mechanics, it is expected that future research into nacre-mimetic materials will be robust.

Nacre organic-mineral bonding also displays high thermal stability. Balmain utilized X-ray diffraction to demonstrate two transformations of the aragonite mineral structure when subjected to heating (using acetic from the giant oyster, *P. maxima*). At 300–400 °C, aragonite transforms to calcite, while at 500–600 °C the calcite transforms to CaO. The organic matrix of nacre displayed high thermal stability, suffering degradation at 550–600 °C [22]. Due to its high temperature resistance, nacre can polymerize with other materials at high temperatures to attain new composites with improved mechanical characteristics and retained bioactivity.

Several studies have shown that nacre can be transformed into hydroxyapatite, the main inorganic component of vertebrate bone and teeth, via reactions in phosphate buffer solutions. Ni and Ratner found a layer of packed hydroxyapatite particles was formed on the surface of nacre via surface reactions in phosphate buffer at room temperature [87]. These surface reactions comprise a dissolution precipitation mechanism in which calcium ions released from the bulk nacre surface enter the phosphate buffer solution and then precipitate as HA on the nacre surface after interacting with free phosphate. X-ray photoelectron spectroscopy (XPS) and secondary ion mass spectroscopy (SIMS) indicated that the mineral phase of the nacre surface had converted from an aragonite phase to an HA phase [72]. Vecchio converted conch and clam shells to HA via a hydrothermal method, demonstrating the formation of dense HA structures on the shell surface at temperatures around 200 °C. When treated shells were then implanted in rat femoral defects for 6 weeks, the surfaces of the implants were covered by newly formed bone without evidence of a fibrous tissue layer. These results indicated the biocompatibility and bioactivity of the surface converted nacre [88]. Kobayashi found that large amounts of HA particles could be formed on nacre when heated to 200 °C in air followed by soaking in simulated body fluid (SBF). In contrast, when heated to 300 °C, above the temperature where destruction of the organic matrix begins, formation of HA particles was retarded, demonstrating the critical role of the organic matrix in HA formation in SBF [89]. Direct conversion of nacre to HA is also observed under high temperature. Lemos discovered a method of converting milled oyster shell into carbonated HA via a hydrothermal method, demonstrating the formation of dense HA structures on the shell surface at temperatures around 200 °C. When treated shells were then implanted in rat femoral defects for 6 weeks, the surfaces of the implants were covered by newly formed bone without evidence of a fibrous tissue layer.

2.3. Composition of nacre

Although composed of more than 95 wt% inorganic aragonite, the organic matrix (5 wt%) is critical not only to the mechanical properties but particularly the bioactivity of nacre [60,92]. The organic matrix, composed of beta – chitin, silk-like proteins, and acidic glycoproteins rich in aspartic acid, forms the framework between the inorganic mineral phase and has been the subject of extensive research [11,60]. Almeida found that the amino acid composition of the water soluble matrix (WSM) in *P. maxima* is mainly composed of glycine and alanine [93]. Balmain J, using Fourier transform infrared spectroscopy (FT-IR), analyzed the mineral and organic phases of *P. maxima*, revealing the presence of amide, amine and carboxyl groups existing at the organic – mineral interface of the nacreous layer. Following decalcification of the sample, the insoluble organic matrix retained amide, amine and carboxyl groups [22]. These studies demonstrated the tight linkage of the matrix proteins and the aragonite phase, which makes separation and purification of the organic components a major challenge. Water extraction is the most widely used, soft extraction method for nacre matrix proteins compared to other methods (acids, resins, EDTA chelation) capable of minimizing loss of molecules; however, this method is still challenging. Therefore, the purification and classification of the WSM remains a challenge and one of the main research goals for nacre.

Bedouet found that the WSM represents approximately 0.24% of the total weight of nacre, with 60% of the WSM made up of low molecular weight molecules (<1 kDa), which nevertheless have potential bioactivity [94]. Lamghari suggested that the WSM has functions similar to bone morphogen protein (BMP) or transforming growth factor – beta (TGF – B), which induce bone growth [95]. Rousseau demonstrated that the WSM had a greater effect on MC3T3-E1 pre-osteoblast cells than dexamethasone, accelerating differentiation and mineralization [3]. Further, the expression of collagen I and the miRNA expression of Runx2 and osteopontin were increased in MC3T3 cells exposed to the low molecular weight WSM. These results indicated the osteogenic potential of the WSM [96]. Millet conducted further testing on several mammalian cell lines to evaluate the bioactivity of nacre. Bone marrow cells supplemented with WSM began to mineralize following 14 days of culture, and MRC5 fibroblasts displayed an early increase in alkaline phosphatase (ALP) activity [97]. Using 2DE and LC-MS/MS, Oliveiri identified four proteins (three gigasin-2 isoforms and a cystatin A2) for the first time in the WSM of C. gigas nacre. These proteins are thought to play a part in bone remodeling.

<table>
<thead>
<tr>
<th>Table 2</th>
</tr>
</thead>
<tbody>
<tr>
<td>Bioactive molecules present in nacre.</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Protein</th>
<th>Function</th>
<th>Ref.</th>
</tr>
</thead>
<tbody>
<tr>
<td>AP7</td>
<td>Modulates matrix assembly, nucleation, and crystal growth</td>
<td>[108–111]</td>
</tr>
<tr>
<td>AP8</td>
<td>Specifically modifies calcite crystal morphology</td>
<td>[112]</td>
</tr>
<tr>
<td>Dermatopontin</td>
<td>Major matrix component, participates in nacre formation</td>
<td>[113]</td>
</tr>
<tr>
<td>Lustrin A</td>
<td>Controls calcium carbonate morphology and packing</td>
<td>[114]</td>
</tr>
<tr>
<td>MRNP34</td>
<td>Methionine-rich protein associated with biocalcification</td>
<td>[115]</td>
</tr>
<tr>
<td>MS17</td>
<td>Important in determining nacre texture</td>
<td>[116,117]</td>
</tr>
<tr>
<td>MS131, MS160</td>
<td>Major components of shell matrix proteins</td>
<td>[118]</td>
</tr>
<tr>
<td>N14, N16, perlpin</td>
<td>Exhibits calcium and chitin binding</td>
<td>[119]</td>
</tr>
<tr>
<td>n16</td>
<td>Biogenic mineral stabilization</td>
<td>[120]</td>
</tr>
<tr>
<td>n16.3</td>
<td>Forms gel-like protein phases to organize single-crystal calcite</td>
<td>[121]</td>
</tr>
<tr>
<td>N19</td>
<td>Negative regulatory role in calcification</td>
<td>[122]</td>
</tr>
<tr>
<td>N40</td>
<td>Facilitates nucleation of aragonite</td>
<td>[123]</td>
</tr>
<tr>
<td>Nautilin-63</td>
<td>Binds chitin, exhibits short collagenous-like domains</td>
<td>[124]</td>
</tr>
<tr>
<td>P10</td>
<td>Plays a role in biomineralization</td>
<td>[125]</td>
</tr>
<tr>
<td>P43</td>
<td>EDTA-insoluble matrix protein</td>
<td>[126]</td>
</tr>
<tr>
<td>P60</td>
<td>Stimulates formation of mineralized nodules</td>
<td>[127]</td>
</tr>
<tr>
<td>Perlinhibin</td>
<td>Crystal growth inhibition in nacre formation</td>
<td>[128]</td>
</tr>
<tr>
<td>Per lucin</td>
<td>Functional C-type lectin</td>
<td>[129]</td>
</tr>
<tr>
<td>Per lustrin</td>
<td>Homologue to N-terminal domain of mammalian insulin-like growth factor binding proteins (IGFBPs)</td>
<td>[130]</td>
</tr>
<tr>
<td>Perl wapin</td>
<td>Growth inhibitor for calcium carbonate crystals</td>
<td>[131]</td>
</tr>
<tr>
<td>PIN23</td>
<td>Key accelerator in control over crystal growth</td>
<td>[132]</td>
</tr>
<tr>
<td>PIN44</td>
<td>Stabilizes magnesium calcite to inhibit aragonite deposition</td>
<td>[133]</td>
</tr>
<tr>
<td>Pf</td>
<td>Binds to aragonite crystals and regulates nacre formation</td>
<td>[134]</td>
</tr>
<tr>
<td>Silkmapin</td>
<td>Shell matrix protein involved in nacre formation</td>
<td>[135]</td>
</tr>
<tr>
<td>Upsaline</td>
<td>Mineral associated protein</td>
<td>[136]</td>
</tr>
</tbody>
</table>
and biocompatibility [98]. Almeida found that the WSM from P. maxima could increase ALP activity of cultured human fetal lung tissue fibroblasts [93]. Chaturvedi studied the function of WSM from the nacre of the marine oyster P. fucata, finding both anti-oxidative and osteogenic differentiation potential [99]. Sud investigated the effects of WSM on mantle cell culture of P. maxima for 11 days, finding that the WSM reduced the global viability of the cultured cells while increasing ALP activity, which could indicate selective responses to the WSM by different mantle cell types [100]. Chaturvedi investigated the osteoblast differentiation activity of WSM from P. fucata. The results demonstrated enhanced ALP activity and gene expression of collagen type -1 and osteocalcin [99]. Nacre was also shown to promote wound healing by enhancing cell adhesion and tissue regeneration of skin fibroblasts, stimulating fibroblast mitosis, collagen deposition, and TIMP-1 expression [101].

At the present, many proteins have been successfully purified and identified (Table 2). Bedouet extracted proteinase inhibitors of papain, bovine cathepsin B, and human cathepsin L from the oyster P. margaritifera [102]. Weiss purified perlucin and perlustrin from the shell of the mollusk H. laevigata (abalone) by ion-exchange chromatography and reverse-phase HPLC after demineralization of the shell in 10% acetic acid [103]. Yano identified the N19 protein, an important negative regulatory protein in pearl oyster calcification [104]. Yano also identified a protein family of s-matrices by cDNA library sequencing in the pearl oyster P. fucata, which possessed similar domains comprising repeat sequences of two or more glycines. The results also suggested that s-matrices were secreted into the shell and participated in providing a framework for calcification [89]. Fang found a novel protein, PIN23, a key component that regulates the crystal growth in nacre by increasing calcium carbonate deposition and the induction of mineral components into the crystalline, aragonite structure of the mature bio-composite [105].

Although most of the identified proteins were from the soluble part of the organic matrix, they are also constitutive component of the insoluble matrix under certain conditions. Bedouet identified two nacre matrix proteins in the WSM via a proteomics approach; however, one of these proteins was also a critical part of the insoluble matrix [106]. It is thus possible that certain proteins might be transformed from soluble to insoluble under certain conditions in the process of nacre formation.

Other proteins have been discovered unrelated to biomineralization or shell formation. For example, perlucin displays a C-type lectin function and perlustrin shows homology to the N-terminal domain of mammalian insulin-like growth factor (IGF) [40,107]. Continued research into the protein content of the nacre organic matrix will hopefully reveal the regulatory mechanisms of osteogenesis promoted by nacre.

2.4. Bioactivity of nacre

Although the organic matrix is quantitatively a minor component of nacre, it is the major active component and controls the formation of nacre [105]. For decades, studies have found that nacre is a bioactive material capable of promoting bone formation in vitro and in vivo. However, the mechanism by which nacre stimulates bone differentiation remains largely unknown. It is possible that signal molecules capable of stimulating the osteogenic pathway in mammalian cells reside in nacre [5]. Marin proposed that the organic matrix of nacre was the source of bioactive molecules controlling biomineralization [133]. In addition to its potential as a bone biomaterial, pearl powder and nacre have found applications in beauty products, anti-aging treatments, weight reduction products, and nutritional supplements (via calcium bioavailability) [134–136].

2.5. Biocompatibility of nacre

Nacre and bone are both biomineralized structures, although their composition and organization differ. The mineral phase of human bone is composed of calcium phosphate in the HA crystal form, whereas nacre is composed of calcium carbonate in the aragonite crystal form [87]. Nacre is capable of attracting and activating osteoblasts and is simultaneously biocompatible and osteoinductive, as demonstrated by Silve C [137]. Specifically, perlustrin, an IGF superfamily member found in vertebrates and capable of affecting bone metabolism either by modulating IGF activity or independent of IGF, has been isolated from mollusks and shown to affect osteogenic differentiation [107]. The promising effects of nacre components on mammalian cells with regards to osteogenesis has led to the hypothesis that a similar regulatory mechanism exists in both vertebrates and mollusks. It is possible that the processes of nacre and bone formation, particularly in terms of biomineralization of a calcium based inorganic matrix within an underlying organic framework, are both derived from a common pathway existing during the earliest stages of life, when most, if not all organisms were ocean-bound. It has thus been suggested that nacre and bone have been inherited from a common biomineralization system [6]. This hypothesis has led to the use of nacre as a bone substitute material.

For bone implants, bone bonding is a key factor, and is important for optimal repair. Adhesive surface coatings are required to provide sites for cell anchoring on metal or ceramic implants; however, nacre is capable forming direct bonding with cells. This bonding is facilitated between nacre and newly formed bone via an interlocking calcium and phosphorous front that forms on the nacre surface, resulting from a combination of cell mediated deposition of new mineral as well as conversion of the nacre surface via dissolution – precipitation reactions similar to those observed in vitro [138]. Zeta potential measurements of the interfacial structure at the bone – nacre interface have been used to analyze the structure of the region. Results showed that the primary role of the organic matrix proteins was biocompatibility with bone [139]. Atlan placed nacre within the epiphysis of sheep and after 10 months observed direct binding of the bone to the nacre implant [138]. The cells of the osteoprogenitor layer appeared to be contacting the implant surface directly without intervening fibrous tissue. Although the reason for this result was unclear, it may be due to the favorable microenvironment around the nacre implant.

2.6. Osteogenic activity of nacre

The main characteristics of an osteogenic material are osteoconduction and osteoinduction. It is generally believed that nacre contains one or more signal molecules capable of stimulating bone formation, making it a promising bone biomaterial. Lopez placed nacre chips on a layer of human osteoblasts in vitro, resulting in formation of a dense osteoid matrix containing foci with mineralized structural features and bone – like structures. In the presence of nacre from the shell of P. maxima, bone formation was attained in cultured human osteoblasts, further suggesting the material’s ability to stimulate bone formation [113]. Green also observed the osteogenic potential of nacre chips of P. maxima, with cultured human bone marrow derived stem cells displaying greater ALP expression than BMP-2 treated controls [140]. Lamghari also suggested that nacre has osteogenic activity via in vitro and in vivo experiments [95,141,142]. Chen found that pearl nanocrystallites were capable of improving the osteogenic ability and promoting the osteogenic differentiation of mouse bone cells (MC3T3-E1) in vitro. Avsannov compared the osteogenic effects of nacre and B-TCP on cocultured human bone cells (HBCs) for four weeks.
Results indicated that nacre promoted osteogenic activity by enhancing ALP, bone sialoprotein (BSP), and osteocalcin (OC) expression [143]. Shen demonstrated that, compared to shell nacre and HA, pearl could stimulate the proliferation of osteoblasts more rapidly [4].

The mechanism by which nacre stimulates bone differentiation remains largely unknown. Although most experiments show that nacre is osteogenic, there have been some contrary results. To investigate the interaction between nacre and fresh bone marrow, Bahar placed nacre particles inside demineralized bone matrix (DBM) cylinders filled with fresh rat femur marrow. The cylinders were then implanted subcutaneously at the thoracic region for 4 weeks. The results showed that natural nacre had no osteoinductive performance [144]. Fricain implanted nacre disks or powder from *P. margaritifera* subcutaneously in rats for 12 weeks; however, results suggested that the nacre was biodegradable and biocompatible but not osteoinductive [145].

2.7. Biodegradability of nacre

Biodegradability of nacre depends on the size, shape, and location of implants and can be limited in bulk nacre. Duplat showed that nacre obtained from *Pinctada* oyster shell was capable of stimulating resorption of the material by differentiated osteoblasts *in vitro*; however, the rate of resorption was consistently lower than that of bone [146]. Berland implanted raw nacre into sheep bone. After nine months post implantation, the results suggested that smooth surfaced nacre undergoes limited biodegradation and becomes microporous [147]. Osteoclast activity on nacre was studied by Duplat. Pure nacre, obtained from *Pinctada* oyster shell, acted as a substrate for cell culture of osteoclast stem cells and mature osteoclasts. The results indicated that osteoclasts differentiated from precursor cells on the nacre substrate had the ability to resorb the material. However, compared to osteoclast resorption of bone, the process of nacre resorption was proved to be limited [146]. Liao observed that macrophages and multinucleated giant cells appeared on the degrading nacre implants, indicating biodegradation [148]. Biodegradability of raw nacre is limited *in vivo* due to its well-ordered microstructure. While unfavorable for bone regeneration, this characteristic could be useful in designing artificial dental materials [7].

2.8. Design and applications of nacre-based biomaterials

Due to its mechanical strength, biocompatibility and bioactivity as well as additional properties including fluorescence ability, nacre can be utilized as a biomaterial in a variety of applications (Fig. 2). Major applications include: (1) implantation of nacre blocks directly as a bone substitute or of bulk composite materials containing nacreous powder, (2) direct injection into bone defects or injection within a bone cement, (3) as a coating for implants and drug delivery vehicles, particularly as a coating to improve the biocompatibility of metal implants, either via physical methods such as electrophoresis or via biological methods in which materials are implanted into the mantle of mollusks and act as irritants, prompting their encapsulation in a pearl coating, (4) as osteogenic additives, particularly incorporation of the WSM into existing biomaterials, and (5) within soft materials and as a naturally occurring nanomaterial, fluorescent material, etc. [149]. Although nacre is naturally compact and hard, replacing part of the calcium carbonate content with other, softer materials could result in a more flexible material while maintaining the nanostructure. Additionally, nacre is a natural nanomaterial that can be used in the field of nanotechnology. Amino acid residues within the tablets could be modified to carry drugs, resulting in novel nanocarriers. Unlike HA, nacre also displays natural fluorescence due to components of the organic phase (such as conchiolin), a minimally explored property which could be utilized for imaging [150,151].

![Fig. 2. Design strategies and applications of nacre-based biomaterials.](image-url)
2.9. Direct material implantation

Nacre powder as a bone graft material has been shown to induce new bone formation in vivo. Powdered nacre implanted in human jaw bone defects was capable of inducing bone regeneration, with newly formed bone closely contacting the nacre implant [6]. Significant osteoblast activity (Fig. 3) was present in the newly formed osteoid border regions, suggesting the osteogenic nature of the nacre implant. Liao implanted nacre granules (<2 mm diameter) into rat femurs, displaying the biocompatibility, biodegradability, and osteoconductivity of nacre. Newly formed bone was formed directly on the nacre surface, with a closely fused interface consisting of a phosphorous rich zone [152]. Lamghari mixed nacre powder (50–150 μm) with autologous blood which was injected between the fifth and sixth lumbar vertebrae in an arthritic rabbit model. After two weeks, nacre was well tolerated by the host tissue and after five weeks endochondral bone formation in the region of the degrading nacre particles was detected [153]. After 11 weeks, solid fusion between transverse processes at the implant site was observed [153].

Bulk nacre displays mechanical properties similar to native bone, leading to its use as a bulk grafting material [7]. Nacre plates, screws, and rods were implanted in the first metatarsus of sheep for two months before resection. Results demonstrated the formation of a phosphate rich layer at the bone/nacre interface, without an interposing fibrous tissue layer. Erosion of the bulk nacre was also observed, forming an irregular surface on which osteoblasts were able to generate new bone. Nacre was thus shown to support a direct interface with bone [138]. Milet placed solid shell nacre within the femur epiphysis of sheep, again demonstrating nacre’s ability to support bone tissue generation within an osteoprogenitor rich cell layer [97]. Solid nacre cylinders (6 × 15 mm) were prepared by Atlan and placed within sheep femoral epiphysis. After 10 months, no foreign body response was observed. Moreover, bone formation occurred, with direct binding of newly formed bone with the nacre surface despite minimal degradation of the bulk nacre material [138]. Camprasse tested nacre dental root implants, with excellent union between implanted nacre and human maxillary bone in vivo [7]. Liao compared nacre cylinders to Titania/HA composite implants, resulting in an improved bone/material interface and greater osteogenic activity in the nacre group [9]. Rousseau placed nacre blocks in the subchondral region of the knee in a sheep model to analyze the intra-articular behavior of nacre. New cartilage formation devoid of inflammation was observed after three months [154]. These results demonstrate the potential of nacre as a biomaterial.

2.10. Compositing with other materials

As a raw material, bulk nacre is limited by the difficulty of fabricating desired shapes as well as the slow degradation rate of large implants. Composites composed of nacre and polymeric...
materials have been developed as an alternative biomaterial for orthopedic tissue regeneration, displaying enhanced degradation as well as good mechanical, osteoconductive, and osteoinductive properties.

Introducing nacre into PLLA can improve the mechanical properties of the resulting scaffolds. Liu successfully fabricated PLLA/nacre and PLLA/vaterite scaffolds using the freeze drying method. As pearl powder content increased, porosity decreased and yield strength increased [155]. Fabrication of 3D scaffolds of PLLA/aragonite pearl powder, PLLA/vaterite pearl powder, and PLLA/nacre powder (Fig. 4) demonstrated no physical difference among the composites, which attained nearly twice the compressive strength and compressive modulus of pure PLLA. Cell culture with rat bone marrow-derived mesenchymal stem cells (rBMSCs) resulted in increased cell proliferation and ALP activity in the PLLA/aragonite and PLLA/nacre groups; however, PLLA/vaterite displayed the opposite effect [156]. Comparison of the degradation behavior in phosphate buffer solution (PBS) of PLLA, PLLA/aragonite, and PLLA/vaterite for 200 days demonstrated no difference in degradation rate between the two composites, both of which degraded more slowly than PLLA alone [157]. Dai also fabricated PLLA/nacre powder composites, resulting in an increase in mineralization and cell proliferation versus PLLA alone [158].

A novel, porous PDLLA/nacre nanocomposite scaffold was fabricated by Xiao. The PDLLA/nacre scaffold exhibited a high porosity (82.5 ± 0.8%) and compressive strength in the range of 4.5 ± 0.25 MPa. In vitro and in vivo experiments showed that cells adhered and proliferated well on the scaffolds, demonstrating their biocompatibility. The scaffolds were capable of repairing critical size segmental bone defects in rabbits [159].

Yang prepared a hybrid composite scaffold composed of PLGA and nanonized pearl powder. The scaffolds displayed degradation and osteoinduction in vitro with MC3T3-E1 cells [160]. Xu developed composite scaffolds of poly(Lactic-co-glycolic acid) (PLGA)/pearl using low temperature deposition manufacturing (LDM). Resulting scaffolds had high porosity and sufficient pore size and mechanical properties for bone tissue engineering. In vitro experiments indicated that the composites displayed better biocompatibility and osteoconductivity than tricalcium phosphate/PLGA scaffolds [161].

Liu added ground nacre powder and carbon nanotubes (CNTs) into UHMWPE, coated with perfluoropolyether (PFPE) to fabricate composites. Wear tests demonstrated increased wear resistance for composites versus bulk UHMWPE. Additionally, addition of nacre increased strength versus the bulk polymer [162].

Meifeng fabricated blends of degradable magnesium alloy and pearl powder. Blended alloys were found to alter the corrosion (degradation) rate of the magnesium, creating a more stable implant [163]. Concurrently, inclusion of pearl powder was capable of increasing osteoblast adhesion and proliferation and increasing cytoocompatibility [163].

PEEK is a relatively new class of high performance engineering plastic. PEEK has been used to develop implantable joints and ligaments. Tao manufactured pearl powder/PEEK composite artificial joints composed of a 9:1 ratio of pearl powder: PEEK with a 3 mm thick pearl powder/PEEK layer and a 100 nm pearl powder layer deposited on the bulk implant via a spraying technique. Experimental results showed that the nanometer pearl powder was evenly dispersed and that the composites attained mechanical properties greater than that of titanium composite materials. Cell culture demonstrated a fourfold increase in cell density on pearl powder/PEEK materials versus pure PEEK [164].

Meng blended pearl powder with PHBV to fabricate films, then treated the films with sodium hydroxide (NaOH) and incubated them in simulated body fluid (SBF) to prepare composite films. Results showed enhanced formation of bone-like apatite. Composing with pearl powder also improved the films' mechanical properties and hydrophilicity [165]. Bai electrospun PHBV/pearl powder blends. Compared to neat PHBV, blended fibers displayed improved mineralization and biocompatibility when cultured with MC3T3 cells [166]. Similarly, Deng fabricated electrospun polypropylene/pearl powder composite monofilaments for pelvic reconstructive mesh, with improved cell proliferation versus polypropylene alone [167].
The above composites have demonstrated the ability of pearl powder and nacre to enhance the mechanical and bioactive properties of bulk polymer materials. However, they require fabrication beforehand and necessitate major surgery for implantation. Therefore, the development of injectable nacreous composites should be considered for in situ polymerization, minimizing the trauma of implantation and simplifying manufacturing.

3. Coating of materials

Nacreous coatings are advantageous due to the robust mechanical properties, high biocompatibility, and the ability of the nacre interface to promote bone growth. Zhu demonstrated that titanium surface coated with nacre could promote the proliferation and differentiation of MC3T3-E1 cells in vitro more effectively than either uncoated or HA coated materials. Cultured cells were larger and had a more spread morphology, possibly due to increased gene expression of transforming growth factor - beta (TGF-beta) 1 [168]. Titanium implants coated with nacre from the fresh water bivalve *H. cumingii* Lea cultured with MG-63 cells also promoted the adhesion, proliferation and differentiation of cells [169]. Guo and Zhou fabricated nacre coatings on titanium alloy (Ti6Al4V) via electrophoresis. When subsequently treated in a phosphate solution, the nacre coating was converted via dissolution-precipitation reaction into plate-like apatite with a hierarchical pore structure which showed bioactivity in vitro [170]. These promising in vitro results should lead to further testing of nacre coatings in vivo.

Biofabrication of nacre coatings via direct implantation of materials into the pearl sac of the mollusk has also been studied and is advantageous due to the high mechanical strength of the natural nacre structure deposited on the surface. Coating thickness can be adjusted not only by time but also by implant material choice. Kwan and Wang investigated the biofabrication of nacreous coatings on different materials, indicating the poly(methyl methacrylate) and high density polyethylene were capable of supporting thick coatings (9.8–76.5 μm) while titanium supported relatively thin coatings (4.4–5.9 μm) [171]. Wang cultivated titanium dental implants in the pearl sacs of freshwater bivalves, resulting in nacre coatings of approximately 200–600 μm in thickness after 45 days comprised of a laminated nacreous layer and a transitional, non-laminated layer (Fig. 5). Results suggested that nacre coated titanium dental implants could be fabricated via this novel biological technique to form biologically-active and degradable surface coatings [172,173].

4. Soluble matrix

Nacre, when implanted in vivo in the bones of dogs, sheep, mice, and humans, induces a biological response that includes integration and osteogenic activity that seems to be activated by a set of proteins present in the nacre water soluble matrix (WSM) and ethanol soluble matrix (ESM).

Part of the matrix forming the organic component of nacre is water soluble, and thus may stimulate cells directly in cell culture. Milet investigated the effect of nacre WSM on mammal bone cells, concluding that fibroblasts, human bone marrow stromal cells, and pre-osteoblasts were recruited into an osteogenic pathway, undergoing complete mineralization [97]. Green demonstrated significant increases in proliferation as well as ALP, OCN, and collagen I-IV expression in bone marrow derived stem cells treated with the WSM of *P. maxima* [140]. Using the WSM as a template, Li proposed a new strategy for clinical management of dental caries. WSM was found to be capable of facilitating HA remineralization on eroded

Fig. 5. Biological coating of titanium cultured in pearl sacs for 45 days. (A) and (B) Nacreous coatings on dental implant screws and flat plates respectively. (C) Longitudinal section of coated screw. (D) Enlarged image of selected area from (C) showing the interface between the coating and screw. Reprinted from [148], with permission from Elsevier.
human enamel. The highly ordered HA had similar composition and structure to natural enamel, displaying ideal smoothness and hardness [149]. Recently, Brion successfully restored the mineralization capability of subchondral osteoarthritic osteoblasts after 7 days of treatment with the ESM of nacre [174]. Future study will continue to demonstrate the effects of WSM and ESM on cell lineage commitment.

5. Conclusion

A highly organized organic – inorganic composite material, nacre not only demonstrates excellent mechanical properties due to its complex architecture, but also displays biocompatibility as well as bioactivity that is absent in HA and other ceramic materials commonly used in orthopedic tissue engineering. Nacre as a bulk material, as a coating, or as part of a composite with polymeric materials can be utilized to develop a wide range of biomaterials capable of promoting bone tissue regeneration while remaining degradable. Nacre powder or the WSM can be used as direct injectable materials or part of cements or other self – setting systems. Nacre also has the potential for modification to serve as a drug carrier or to create novel materials demonstrating improved physical properties versus bulk nacre (i.e. accelerated degradation, enhanced elasticity, and improved molding/processing). Similar to HA and calcium phosphate, nacre can also be utilized in three-dimensional printing as a reinforcement material within hard polymers or soft hydrogels. Although currently relatively unknown compared to HA and calcium phosphates, future research will continue to demonstrate the superior bioactivity and novel properties of nacre for orthopedic tissue regeneration, leading to a new class of materials and composites based on this biologically derived product.

Acknowledgement

This work was supported in part by National Institutes of Health awards (CA182670, HL118498) and National Science Foundation (NSF) award (CMMI 1537008).

References


M. Xu, Y. Li, H. Suo, Y. Yan, L. Liu, Q. Wang, Y. Ge, Y. Xu, Fabricating a pearl/PLGA composite scaffold by the low-temperature deposition manufacturing technique for bone tissue engineering, Biofabrication 2 (2010) 025002.


